

# Gonadotropin-releasing hormone and adipokinetic hormone signaling systems share a common evolutionary origin

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Gonadotropin-releasing hormone (GnRH) is a critical and central hormone that regulates vertebrate reproduction. The high conservation of GnRH signaling within the chordates (deuterostomians) raises the important question as to whether its appearance might date back prior to the divergence of protostomian and deuterostomian lineages, about 700 million years ago. This leads to several important questions regarding the evolution of the GnRH family. Has GnRH been retained in most protostomian lineages? And was regulation of reproduction already a function of ancestral GnRH? The first question can undoubtedly be answered affirmatively since several GnRH-like sequences have been found in wide variety of protostomian and deuterostomian phyla. However, based on their different primary functions in different phyla – which implies a less unanimous answer on the second question – consistency in the nomenclature of this peptide family has been lost. A comparative and phylogenetic approach shows that the ecdysozoan adipokinetic hormones (AKHs), lophotrochozoan GnRHs and chordate GnRHs are structurally related and suggests that they all originate from a common ancestor. This review supports the view that the AKH–GnRH signaling system probably arose very early in metazoan evolution, prior to the divergence of protostomians and deuterostomians.

**Keywords:** gonadotropin-releasing hormone, adipokinetic hormone, neuropeptide, G protein-coupled receptor, reproduction, metabolism, evolution

## INTRODUCTION

Gonadotropin-releasing hormone (GnRH) is well known for its control of vertebrate reproduction, via the pituitary, where it regulates the synthesis and release of the gonadotropins luteinizing hormone (LH) and follicle-stimulating hormone (FSH; Kah et al., 2007). Its mammalian structure was originally thought to be unique until it became apparent that diverse forms of this peptide hormone exist in vertebrates. At present, the chordate GnRH family consists of 25 members: 16 variants have been found in vertebrates (Morgan and Millar, 2004; Guilgur et al., 2006; Kavanaugh et al., 2008; Kim et al., 2011), and nine in tunicates (Adams et al., 2003). Some vertebrates, like chickens, express two and some teleost fish even three different forms of GnRH (Millar, 2005), which is the result of gene duplications, diversifications, and losses (Stewart et al., 2009). Recent studies demonstrate by genome synteny analyses that the GnRH family most likely comprises three paralogous genes (GnRH1, GnRH2, and GnRH3) resulting from a genome/gene duplication event affecting the common ancestor of lamprey and gnathostome isoforms. An additional group of GnRH4 isoforms (consisting of lamprey GnRH-I and GnRH-III) is present only in lampreys without any counterpart in the gnathostome lineage (Kim et al., 2011; Tostivint, 2011). All these chordate GnRHs signal through G protein-coupled receptors (GPCRs), which consist of seven-transmembrane (7TM) helices and signal through Gαq with Ca<sup>2+</sup> as second messenger. GnRH

receptors have been reported in the cephalochordate *Amphioxus*, in tunicates, agnathans, and in various vertebrates (Kah et al., 2007; Tello and Sherwood, 2009).

The existence of a putative functional equivalent of this peptide hormone and its receptor in protostomian invertebrates has been a matter of debate. Because the basic concepts of sexual reproduction are similar in most animal phyla, it is tempting to hypothesize that the regulation of reproduction must have been highly conserved and may date back to the common ancestor of protostomians and deuterostomians. The first reported protostomian GnRH sequence has been reported in the mollusk *Octopus vulgaris* (Iwakoshi et al., 2002). Until recently, GnRH-related peptides were only established in two protostomian phyla belonging to the Lophotrochozoa: the mollusks (*Sepia officinalis*, Di Cristo et al., 2008b; *Aplysia californica*, Zhang et al., 2000; and *Lottia gigantea*, Tsai and Zhang, 2008) and annelids (*Capitella teleta*, Tsai and Zhang, 2008; and *Helobdella robusta*, Roch et al., 2011).

## IS GnRH ALSO PRESENT IN ECDYSOZOANS?

Recently, Tsai and Zhang (2008) reported that chordate and protostomian GnRHs likely share a common ancestor. They proposed that GnRH has been preserved in lophotrochozoans (annelids and mollusks), but lost in the ecdysozoan lineage (nematodes and arthropods; Tsai and Zhang, 2008). According to their view, GnRHR-like receptors have been retained in Ecdysozoa but they

have recruited other ligands to bind. The ligand of the GnRH receptor of the insects *Drosophila melanogaster* and *Bombyx mori* has been identified as adipokinetic hormone (AKH; Staubli et al., 2002). AKH is an insect neurohormone that is synthesized in the corpora cardiaca (CC; O'Shea and Rayne, 1992), a neurohemal organ which is considered as the functional equivalent of the vertebrate pituitary gland. AKH is involved in mobilizing energy-containing substances such as sugar and lipids from the fat body during flight and locomotion (Gade, 2004).

A breakthrough into the common evolution of AKH and GnRH came with the demonstration of an AKH–GnRH-like neuropeptide in the nematode *Caenorhabditis elegans* (Ce-AKH–GnRH), which is reminiscent of both insect AKHs and of GnRHs from tunicates and higher vertebrates (Lindemans et al., 2009). Although it has a small size and, at first sight, simple anatomy containing only 302 neurons, this small soil nematode has already repeatedly proven its value in neurobiological and endocrinological research (Janssen et al., 2010). The *C. elegans* AKH–GnRH neuropeptide has been found by a bioinformatics approach based on the Prosite pattern that characterizes the AKH family. The amino acid sequence of the novel *C. elegans* neuropeptide (pQMTFT-DQWT) is more similar to AKH than to GnRH (Figure 1). Ce-AKH–GnRH activates in a dose-dependent way a *C. elegans* GPCR (Lindemans et al., 2009) that is homologous to the human GnRH receptor and has been designated as the *C. elegans* GnRH receptor (Ce-GnRHR; Vadakkadath et al., 2006). This receptor signals through Gαq, analogous to most AKH and GnRH receptors. EST libraries available on the web (<http://www.nematode.net/BLAST/>) and a NCBI BLAST search revealed orthologous AKH–GnRH-like precursor sequences in parasitic nematodes (Figure 1). All these nematode AKH–GnRH sequences activate the Ce-GnRHR in a dose-dependent way (Lindemans et al., 2009), demonstrating that the AKH–GnRH neuropeptide is widespread throughout the phylum of nematodes.

### GnRH AT THE CHORDATE ROOT

Non-vertebrate chordates, like *Amphioxus*, provide very valuable information about GnRH signaling at the basis of the chordate lineage. The *Amphioxus* genome encodes, as a result of gene duplications, four GnRH receptors. GnRHR1 and 2 have a higher degree of conservation with vertebrate orthologs and are activated by vertebrate GnRHs, whereas GnRHR3 and 4 rather cluster with protostomian GnRH receptors (Roch et al., 2011). An additional strong argument for the common origin of AKH and GnRH came with the demonstration of insect AKH and mollusk GnRH activating the *Amphioxus* GnRHR3 in a dose-dependent way, even with a higher potency than two chordate GnRH sequences (GnRHR4 was non-responsive; Tello and Sherwood, 2009). This indicates that *Amphioxus* receptors might provide an explanation for the evolutionary distinct protostomian and deuterostomian GnRH signaling systems.

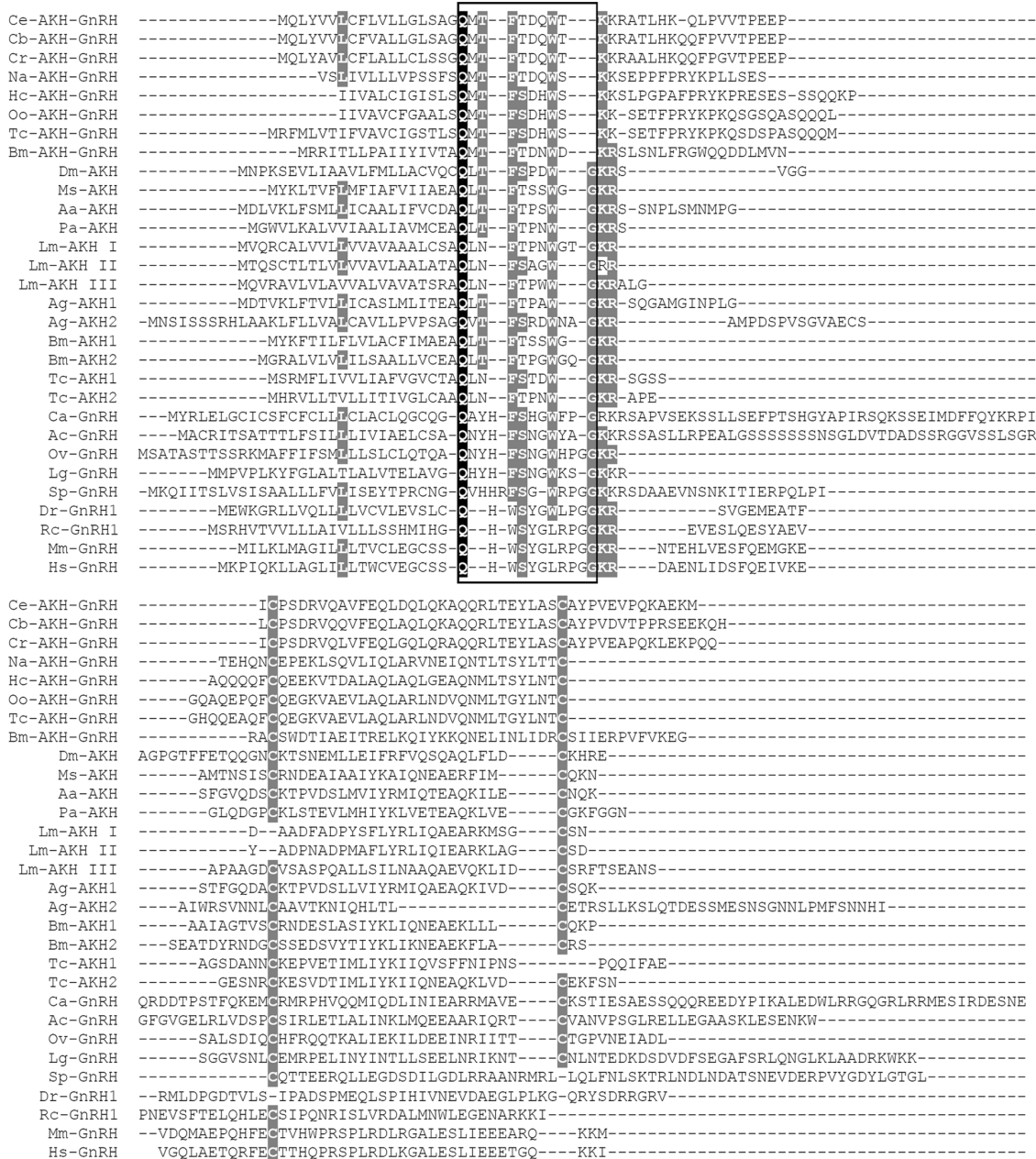
### THE AKH–GnRH SUPERFAMILY

Chordate GnRH is a short, highly conserved decapeptide hormone with four identical residues (positions 1, 4, 9, and 10) of the 10 (Gorbman and Sower, 2003; Kah et al., 2007). Lophotrochozoan GnRHs count usually 11 or 12 residues and ecdysozoan AKHs 9 or

10 (Figure 1). All known AKH and GnRH peptides start with an N-terminal pyroglutamate residue and all processed peptides are amidated except the nematode ones. A plausible hypothesis is that a non-amidated AKH is the most ancient form and that amidation arose later in evolution. However, it is also possible that amidation might have been lost later in evolution. Non-amidated AKH is also found in the painted lady butterfly *Vanessa cardui* (Kollisch et al., 2000) and in the fall armyworm *Spodoptera frugiperda* (Abdel-Latif and Hoffmann, 2007) both belonging to the Lepidoptera. The F(S/T) signature in the middle of the AKH–GnRH peptide sequences is common to all protostomian peptides and, remarkably, also in the echinoderm sea urchin (deuterostomian) *Strongylocentrotus purpuratus*, which resembles more the protostomian GnRHs from annelids and mollusks than the chordate GnRHs that are characterized by a middle WS motif. Further on in the peptide sequence, a tryptophan residue has been well conserved in the protostomian lineage, in the more ancient deuterostomians and also in the ancient chicken GnRH2 (QHWSHGWPY-NH<sub>2</sub>), but has been lost in the amphibians. Moreover, there are striking overall structural similarities between AKH and GnRH precursors (Lindemans et al., 2009): the GnRH or AKH peptide sequence is located immediately after the signal peptide and is flanked at the C-terminus by a basic cleavage site. The C-terminal flanking peptides contain two cysteine residues in all protostomian AKH–GnRH prepropeptides, except for locust AKH-I and II. Associated peptides in the chordate precursors have one or no Cys residue. All these findings provide strong evidence for a common origin of AKHs and GnRHs (Figure 2). The nematode and arthropod (ecdysozoans) AKH–GnRH-like-precursor seems to have diverged substantially from chordate GnRH during the course of evolution. This finding in combination with the rapid divergence of short neuropeptide sequences hampers the identification of homologous genes in the different phyla by comparative genomic tools (e.g., BLAST).

Another group of peptides that are structurally related to AKHs are the corazonins. The first member of this class of peptides – present in insects and crustaceans – was identified from the CC of the cockroach *Periplaneta americana* (Veenstra, 1989). Corazonins have very varying functions in different species, but no generalized role. In cockroaches, they have cardiostimulatory effects (Veenstra, 1989), while in locusts they cause increased pigmentation (Hua et al., 2000) and in silk worms a decreased silk production (Tanaka et al., 2002).

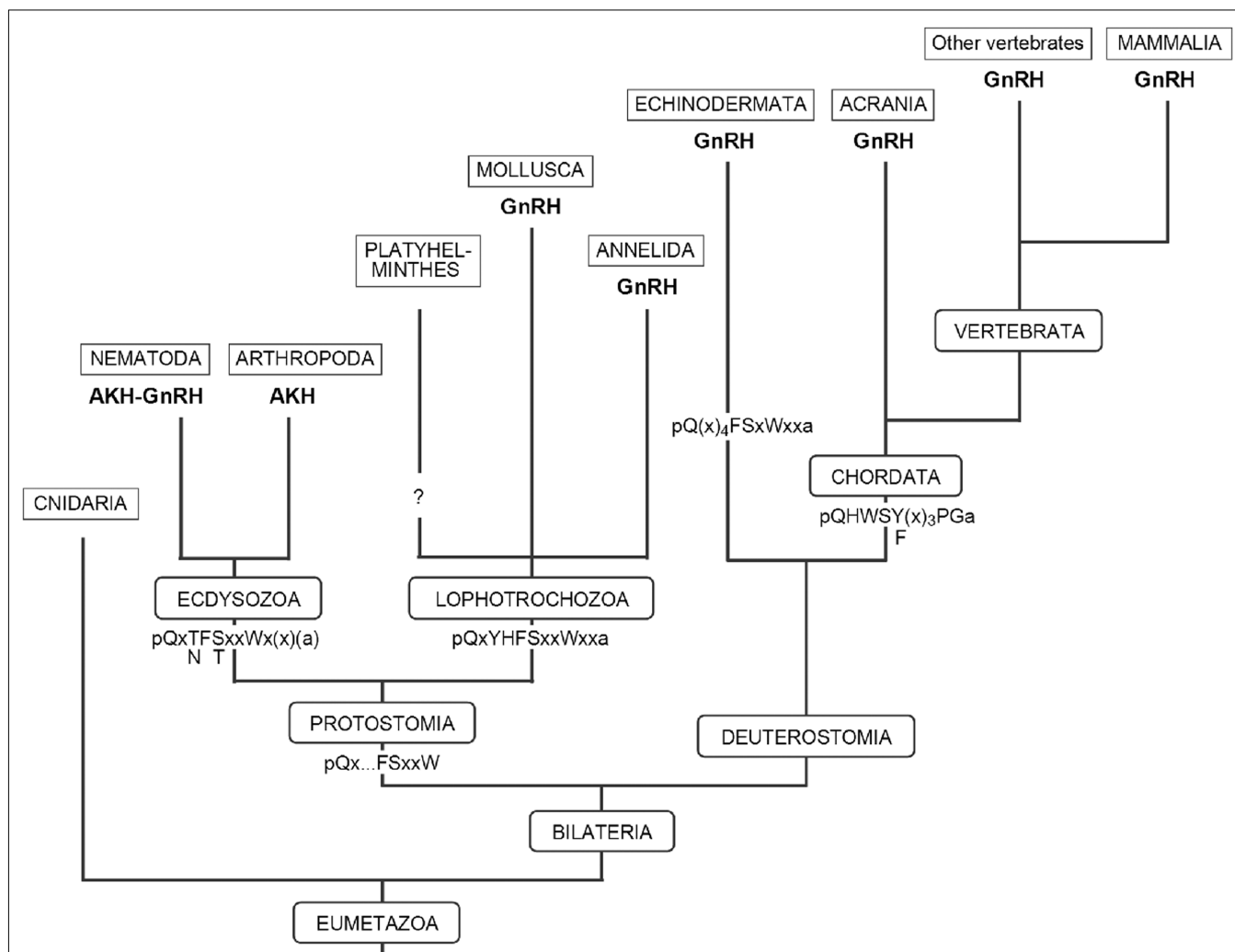
Very recent evidence for the close relationship between both insect neuropeptide groups came from the discovery of an AKH/corazonin-related peptide (ACP; pQVTFSRDWNA-NH<sub>2</sub>) in the mosquito *Anopheles gambiae* (Ag-AKH2 in Figure 1; Hansen et al., 2010). Apart from two carboxyterminal amino acids, this *Anopheles* sequence is completely identical to hypertrehalosaemic hormone of *Locusta migratoria* (Lm-HrTH), another AKH-like neuropeptide (pQVTFSRDWSP-NH<sub>2</sub>; Siebert, 1999). In contrast to Lm-AKH-I, II, and III, which are present in the glandular lobe of the CC, Lm-HrTH is present in the storage lobe of the CC, indicating that this neuropeptide is synthesized in the brain. For this reason and also since there is 36–59% sequence identity between Lm-HrTH and the three “classic” AKHs, but 78% identity with *Drosophila* AKH, it has been suggested before that



**FIGURE 1 | Amino acid sequence alignment of AKH and GnRH peptide precursors of different protostomian and deuterostomian representatives.** Alignment was generated by using the program Vector

NTI. Identical amino acids are highlighted in black and similar amino acids in gray. The concerned peptides are framed and cysteine residues moreover indicated with an arrow. Note that the sequences coming from nematodes other than the *Caenorhabditis* species and *Brugia malayi* are ESTs. Accession numbers in EMBL/GenBank; nematodes: *Caenorhabditis elegans*: Ce-AKH-GnRH (AAC26928), *Caenorhabditis briggsae*: Cb-AKH-GnRH (CBG19970), *Caenorhabditis remanei*: Cr-AKH-GnRH (cr01.sctg44.wum.78), *Necator americanus*: Na-AKH-GnRH (EST BU666660), *Haemonchus contortus*: Hc-AKH-GnRH (EST BF186675), *Ostertagia ostertagi*: Oo-AKH-GnRH (EST BQ625695), *Teladorsagia circumcincta*: Tc-AKH-GnRH (EST CB043182), *Brugia malayi*:

Bm-AKH-GnRH (XP\_00189879), Insects: *Drosophila melanogaster*: Dm-AKH (P61855), *Manduca sexta*: Ms-AKH (P67788), *Aedes aegypti*: Aa-AKH (XP\_001655817), *Periplaneta americana*: Pa-AKH (AAV41425), *Locusta migratoria*: Lm-AKH I (P55319), Lm-AKH II (P08379), Lm-AKH III (P19872), *Anopheles gambiae*: Ag-AKH1 (XP\_001689190), Ag-AKH2 (XP\_563757), *Bombyx mori*: Bm-AKH1 (BAG50369), Bm-AKH2 (NP\_001124365.1), *Tribolium castaneum*: Tc-AKH-1 (NP\_001107797), Tc-AKH2 (NP\_001107818), Annelids: *Capitella* sp., (polychaete annelid), Ca-GnRH (EY629959), Mollusks: *Aplysia californica*: Ac-GnRH (ABW82703), *Octopus vulgaris*: Ov-GnRH (13), *Lottia gigantea*, Lg-GnRH (FC805608), Echinoderms: *Strongylocentrotus purpuratus*, Sp-GnRH (XP\_800179), Vertebrates: *Danio rerio*: Dr-GnRH1 (AAL99294; bone fish), *Rana catesbeiana*: Rc-GnRH1 (AAL05972; amphibian), *Mus musculus*: Mm-GnRH (NP\_032171), and *Homo sapiens*: Hs-GnRH (NP\_000816).



**FIGURE 2 | Diagram showing the evolution of GnRH, AKH, and AKH-GnRH within the Eumetazoa.** Conserved amino acid sequences of the AKH-GnRH neuropeptides are depicted at the roots of the clades. Note that in the Cnidaria and the Platyhelminthes, neither GnRH- nor AKH-like neuropeptides have been found so far.

hypertrehalosaemic hormone might be the most ancient AKH molecule (Siegert, 1999). So far, the precursor sequence of Lm-HrTH is unknown. However, similar hypertrehalosemic peptides and their precursor genes have been found in the genome of other insects, including dipterans (*A. gambiae* (Kaufmann and Brown, 2006, 2008; Hansen et al., 2010), lepidopterans (*B. mori*; Zhu et al., 2009), and coleopterans (*Tribolium castaneum*; Li et al., 2008; Figure 1).

#### ARE PROTOSTOMIAN GnRHs INVOLVED IN REPRODUCTION?

Based on its expression pattern in the brain and ovary, it has been proposed that mollusk GnRH plays a role in reproduction (Di Cristo et al., 2008a,b). A GnRH receptor has been reported in the mollusks *O. vulgaris* (Kanda et al., 2006) and *Crassostrea gigas*, where qRT-PCR showed a specific expression of Cg-GnRHR in both male and female gonads during the reproductive cycle (Rodet et al., 2005), demonstrating for the first time the plausible involvement of a GnRH receptor orthologue in the control of

reproduction in a protostomian invertebrate. Ecdysozoan AKH-GnRH involvement in reproduction was for the first time provided by RNAi experiments of the AKH-GnRH precursor and the GnRH receptor in *C. elegans*. It is clear that Ce-GnRHR and Ce-AKH-GnRH affect the same pathway, as their knock-downs yielded the same phenotype: a delay in timing of the egg-laying process as well as a decrease in the total amount of progeny (Lindemans et al., 2009).

Insect AKH, especially involved in energy mobilizing during flight and locomotion, is so far not linked with a role related to reproduction. AKH deficient fruit flies seem to display normal reproductive capabilities, but, to our knowledge, an investigation toward delayed egg-laying has not been performed yet. It is interesting to note that in the fall armyworm *S. frugiperda* AKH mRNAs occur in ovaries, midgut, fat body, accessory glands, and muscle tissues, suggesting that *akh* genes may play a role in the regulation of oocyte maturation (Abdel-Latif and Hoffmann, 2007). Also in *A. gambiae* females, AKH receptor transcripts are abundant in the

head, thorax, dorsal, and ventral abdomen walls to which most of the fat body is attached and in ovaries as well (Kaufmann and Brown, 2006). And in locusts, the ancient Lm-HrTH has no biological activity usually associated with the typical AKHs (Siebert, 1999). Its biological role remains to be established, and it would be interesting to determine if it is involved in reproduction.

## CONCLUSION

Gonadotropin-releasing hormone is a critical neurohormone involved in the regulation of reproduction in vertebrates and has extensively been studied in this clade. However, the time that GnRH signaling was considered to be unique to chordates has undoubtedly passed. At the moment, GnRH-like peptide sequences have been found in a wide variety of deuterostomian and protostomian phyla including indications of GnRH-like material in the cnidarian lineage (Ancil, 2000; Twan et al., 2006). However,

based on their primary functions in different phyla, consistency in the nomenclature of this peptide superfamily has been lost. It is clear that a comparative and phylogenetic approach, also thoroughly reviewed in (Roch et al., 2011) opens the view that the GnRH superfamily is composed of the ecdysozoan AKHs, lophotrozoan GnRHs, and (proto) chordate GnRHs. They all originate from a common ancestor that may represent one of the most ancient hormones to persist through metazoan evolution.

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